**Bee Diversity in Lowland Puget Sound Marginal Lands**

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ABSTRACT

Bee diversity was assessed at 3 marginal, peri-urban sites in the middle Puget Sound over a period of 7 years. A standardized deployment of pan and “blue vane” traps and opportunistic sweep netting were used on a monthly basis, ultimately producing a total of 25,400 specimens from 155 morphogroups from at least 141 species. Chao richness estimation suggests that the overall bee gamma richness could be as high as 182.5, with the greatest contributions from the genera *Andrena* (Andrenidae,) *Osmia* (Megachilidae,) and the “*Evalaeus/Hemihalictus*” complex (Halictidae.) We found <> introduced species, <> County and <> state records and the apparent absence of the threatened *Bombus occidentalis*, despite system trapping in previously known habitat. The data will provide part of a bee diversity baseline for the region.

INTRODUCTION

The bee diversity of western Washington has remained largely undescribed until recently. Bloom et al. conducted an extensive study of bee diversity in the region, considering farm, garden, and parkland habitat (<Boom et al>.) Prior to this, the only major published assessment of bee diversity to date has been Griswold and Tepedino (<1980>) who cataloged bees of the Columbia Basin, ecologically a region very distinct from the maritime Puget Sound zone west of the Cascade Range.

In 2014 The Common Acre, a private non-profit group supporting science and the arts, obtained funding to attempt vegetative restoration of two areas 1) power corridors of the local power utility, Seattle City Light, and 2) a buffer region surrounding the Port of Seattle’s SeaTac Airport. The central purpose of the foundational study, was, therefore, restoration monitoring. It was decided after implementation to measure the success of the restoration effort in part by assessing proximal native bee diversity in a before/after context. A sister site at Boeing Paine Field outside of Everett, WA was added to the restoration and bee monitoring effort in 2018. These sites share habitat characteristics (see Methods) and are distinct from and complimentary to the study areas used in Bloom et al. (2022).

In 2014, TCA engaged the senior author to direct the bee survey work and the second author was soon after employed to join the team. These personnel had recently been involved, as a separate project, in surveying the Puget Sound and some peripheral areas for *Bombus occidentalis*, populations of which have been rapidly declining since about 2000 [do we have a published reference for this?]. The non-profit BeeSearch was founded as a vehicle for the latter work. Upon joining the TCA restoration effort, the authors quickly saw, in addition to addressing restoration monitoring responsibilities, the opportunity to begin a much-needed intensive, long-term, stand-alone bee diversity assessment of the targeted land. The Common Acre initially engaged BeeSearch as a contractor, but the two organizations later functioned under a joint operating agreement with cataloging bee diversity as a goal. In 2017, the vegetation restoration efforts were discontinued and all support was put toward bee monitoring until final data collection in 2020.

The initial focus of the study was to simply document the bee species richness in the study areas. With the addition of the third author for the purpose of deepening our analyses, the study was broadened to include comparison between seasons, between sites, and more sophisticated considerations of richness.

METHODS

Study sites.

Western Washinton; need to start broader, then describe the individual sites.

1. Port of Seattle (POS, SeaTac Airport.) Used 2014-2020. This site is approximately 3 km2 south of the main runways and extending to the southern limit of Port jurisdiction. This land functions as a security buffer zone around the airport. Through the middle of the last century it was largely used as a private golf course, remnant components of which are recognizable, e.g. “sand trap” areas, hardscape around the former clubhouse. We categorize it as “marginal” because of the broad and varied disturbances historically incurred, proximity to intense development, and present state of low- to no management. The vegetation is varied, with a riparian-like zone along a drainage ditch and several plots of attempted past revegetation. The vegetation is mainly “weedy” with notable border hedges of wild Himalayan blackberry, *Rubus befrons*, patches of knapweed, *Centaurea* sp., wild mustard , *Brassica* spp., and edge patches of Big Leaf Maple, Acer macrophyllum. Willow, *Salix* spp. and a mix of tree and shrubs characteristic of Puget Sound lowland wild areas. Open areas with low perennial or annual weedy plants provided the main collecting areas. Trapping stations in this site were separated by at most a few hundred meters. Notably, within 300 m and 1 km of trapping stations were two honey bee (*Apis mellifera*) apiaries with up to 10 functioning hives.
2. Seattle City Light (SCL, power corridor buffer.) Approximatley 10 km SSE of Seattle, King County, WA. Used 2014-2016. This site consisted initially of 4 trap stations located approximately 2 km apart along an E-W transect along the power corridor beginning in the Rainier Beach neighborhood of south Seattle in a semi-managed meadow-like habitat. A second station occupied semi-boggy slope descending toward the Interstate 5 corridor. A third station was formed west of I-5 within the bounds of a weedy SCL equipment storage pad and immediately North of the Duwamish Hill Preserve. A fourth station was established in the Boulevard Park neighborhoos along the banks of the Duwamish River to the northwest of station 3 between a major road and an SCL power transfer station. A fifth station was added in 2016 approximately 100 m to the southeast of station 1 and in essentially identical habitat. In aggregate, “marginal” also describes this complex site for the reasons stated above.
3. Boeing Paine Field (BPF, airport and maintenance facility.) Used 2018-2020. Approximately 10 km SW of Everett, Snohomish County, WA. This site, trapped from 2018-2020, consisted of 5 stations with separations of 50-500 meters. Habitat varied from low-maintenance lawn/meadow to riparian creek/pond, all surrounded by a narrow corridor of red alder (Alnus rubra)-dominated forest beyond which is suburban development and/or airport parking lots and runways. This site is approximately 1 km from the waters of Puget Sound and experiences, compared to the other sites, significant maritime influence. The varied disturbance and non-natural nature of the land again qualifies this site as “marginal.”

<<Descriptive site aspect photos, a few close-ups of primary resources (Figs.)>>

Field Collecting:

Trap stations consisted of uniform linear arrays of 15 6-inch plastic cereal bowls (Solo) placed in 5 clusters of 3 each white, blue, and yellow. These large bowls were replaced in 2016 and subsequently with identically-painted 4 oz. “mini-bowls” with, according to literature ( ) equivalent attractiveness to bees. The yellow and blue paint used was () as used in other bee surveys (). The bowl arrays were accompanied by 3 equally-spaced “blue vane” traps (BanfieldBio, Woodinville, WA) with original fluorescent yellow collecting jars. Thus, we designate this 15+3 configuration as our basic functional trapping unit. A significant variation was employed at BPF, where only a single Blue Vane Trap per 15-bowl array was used. This was a conservation measure in consideration of the potential presence of the threatened Bombus occidentalis, the last know population of which in the Puget sound was within flight range of BPF. <Rough calculations based on number of specimens collected in bowl vs. Blue Vane traps and consideration of such comparisons in the literature ( ) suggests that this variation had approximately 0.6 X the power of the full trap unit.> This was a conservation measure in consideration of the potential presence of the threatened Bombus occidentalis, the last known population of which in the Puget sound was within flight range of BPF. In several of the early years of the survey, traps placed in high vegetation were elevated on wooden stakes with specially designed platforms in an attempt to increase visibility of the bowl. Likewise, in a few instances, Blue Vane traps were suspended from metal rods with a terminal hook. These elevated methods were found not to increase trap catch substantially in our sites, added substantially to deployment effort, were not widely used, and were abandoned in the later years of the study.

Traps were filled with a catch fluid of water containing 5-10 drops of unscented dish detergent. Trap placement was done on a monthly bases from April through September in most years, with collection date sensitive to weather conditions. Trap arrays were placed for a single 24-hour period starting at approximately 8:00-10:00.

Trap collecting was supplemented with net collecting on an opportunistic basis, as described elsewhere (Turley et al. 2024). The method used was most often sweeping with a heavy 15-inch sweep net bag and the standard measurement of effort consisting of 100 sweeps, although this frequently varied according to the extent of substrate.

<Comparative collecting effort between years and between sites, with the main variable being number of trap placement days and the secondary variable proportion of basic trapping unit, supplemented by net collecting, is diagrammed in Table 1.> move to Results.

Specimens were collected from traps by pouring the trap fluid through a fine strainer and then placing directly into pre-prepared and labeled vials filled with 70% ethanol. Trap specimens were segregated by Date/site/station. Net-collected specimens were dispatched with ethyl acetate and then placed in paper and blotter paper “layers” for temporary storage (). Net collected specimens received a number that corresponded to nearest station or stations if in between.

<< THIS SECTION NOT NECESSARY???

Trap function notes

Uniformity of catch

Large vs. small bowls (comparative data?) <Need a literature reference (Droege?) comparing small vs. large bowls or else our own data, analysed, under Results. >

Small bowls used after 2015(?)

Source of bowls

Stands (comparative data?); vs ground placement

Backpack sprayer for bowl filling (Fig., photo)>>

*Lab procedures*

Pickling vs. pinning

Preserving most common, most easily identifiable

Drying of alcohol specimens for pinning <ref. Droege?>.

Data & Specimen filing

Data collection conventions. Dates, combining trap catches, etc. <Combined trap catches for each station, i.e. did not distinguish blue-vane from bowl catch.)

Identification. Process: grouping, high to lower level. Resources at USDA, ARS, PIRC.

Specimen & data repositories.

% in alcohol

location of specimens

*Data Recording & Specimen Processing*

In lab, specimens were removed from alcohol and then grouped taxonomically to the lowest level allowing certainty of identification to species or morphospecies. Data with tentative taxonomic identifications was recorded into a central data base, one record per specimen. Of each included taxon and sex a representative series was then dried in a small tea strainer capsule with a hair drier heat source as described by <Droege et al.> Once so prepared, specimens were then pinned or pointed and set into collection event series. Pre-prepared labels were then applied, including for each specimen a unique project-specific accession number. The number of leftover specimens from each taxon were stored in an archival glass vial which was labeled with the complete collection information. Data entry reflected all pertinent collection information plus putative identification for all specimens. Data was recorded for extra specimens as a separate record with the number of specimens represented. Storage was in standard Schmitt boxes or, once identified, in regulation insect cabinets with preservative.

*Specimen Identification*

Specimens were identified using a variety of resources, including published taxonomic papers, semi-technical guides, on line Discover Life keys, and <>. Professional assistance was sought from a number of specialists by sending them representative samples. We also made a number of personal visits to the USDA Pollinating Insects/Utah State University Bee Collection in Logan, UT to use the extensive collection there and consult directly with resident specialists. Similar help was also sought at the Bohart Museum of Entomology, University of California, Davis, CA. Specimens that eluded species identity were assigned mophospecies numbers that were unique across the collection. Separate numbers were assigned to females and males as correspondence was usually also uncertain.

*Stastistical Analysis*

Riley’s descriptions I will add any data methods once we’ve determined which analyses and figures we’re presenting.

Data & Specimen deposition

Data will be made available to the Washington Bee Atlas project and representative specimens will ultimately reside in the collection of Washington State University. Additionally, all data have been deposited at Dryad (), and all analyses are archived on GitHub ().

RESULTS

Simple findings

Across the three sites and seven years of the project we collected a total of 25,441 bee specimens. Of these specimens, 25,017 (98.33%) were identified to the species level, representing 118 species from five families and 24 genera. Of the remaining 424 specimens, 376 (1.48%) were identified to morphospecies, representing potentially 35 additional species within the genera: Nomada, Osmia, Sphecodes, and Triepeolus. The remaining 48 specimens (0.19%) were damaged, hindering species-level identification, however, these specimens were confidently identified to genus.

Our species accumulation curves did not reach asymptotes, suggesting that the 118 confirmed species is likely an underestimate of the true richness across the three sites (Figure X). Chao richness estimation revealed the total richness across all sites and years as 139 species (using trap and net collected records). Using only trap records, Chao richness was estimated at 124 species. Chao richness for the individual sites, POS, SCL, and BPF was estimated using trap records only at 115, 92, and 80 species, respectively.

A graph of different sites

Description automatically generated with medium confidence

**Species accumulation curves** for all sites with blue vane trap and net sweeping (black), all sites with only blue vane traps (maroon), Port of Seattle (yellow), Seattle City Lights (blue), and Boeing Plane Field (green). Collection effort (number of sampling sites) was derived from permutation resampling of the subsites within each site for each year.

We found <This is not in the data> introduced species accounting for <>% of the overall richness and amounting to <> specimens. These are listed in Table 2. *Also put in Table 3?*

Species which were represented by a single specimen (“singletons”) have a disproportionate influence on some analyses, such as Chao richness and may have special significance regarding collecting technique, conservation, and biogeography (). <Twenty> of our recovered species (<16.95%> of total richness) were represented by singletons, which are listed in Table??. In a similar vein, super-abundant species can reflect importantly on community structure and we therefore list in Table 3 the top 10 species by specimen representation. Breadth of distribution can be important in community considerations, so we also list in table 3 those species that were collected in all 3 of our sites. As a comment on collecting technique and the importance of net collecting to supplement traps, we found <11> species collected only by net, representing <9.32%> of total richness. See [GitHub/SAC\_and\_Chao](https://github.com/andersonrm/BeeSearch/blob/main/analyses/SAC_and_Chao.md) for these tables generated individually. Not sure how we can make one sensible table out of all of this information.

Males are often underrepresented in collections for various reasons; we finally list in Table 3 species for which we collected no males by either traps or netting. Is this something we really need?

We collected a total of <575> specimens of *Apis mellifera*, representing 2.26 % (575/25441). Of the 575 *A. mellifera* specimens, 213 (37.04 %) were collected by net with the remainder in traps, mostly Blue Vane Traps.

This study did not focus on documenting in detail the differences in trap function regarding bowl vs. Blue Vane or between various bowl colors. Anecdotally, however, we can report that Blue Vane traps excelled in collecting large bodied species and a greater diversity than bowl traps, yet the two trap types were complementary in function. What is this? I don’t have a distinction between traps in the data.

Of the 24 genera found, overall, *Lasioglossum* was the most speciose (21 species), while *Halictus* was the most abundant, representing 47.01% of all records (Figure Xa). However, two sites, SCL and BPF did not follow the overall abundance pattern. In these sites, *Bombus* was by far the most abundant genus, with 26.10% of records in SCL, and 42.80% in BPF (Figure Xc,d).

A graph of different colored lines

Description automatically generated with medium confidence

**Proportional abundance and unique species.** A) All sites collectively, B) POS, C) SCL, and D) BPF. Data are from trap collected records and exclude all net caught records. Data are pooled across years.

Alternatively:

A group of colorful bars with black text

Description automatically generated with medium confidence

**Proportional abundance and unique species for all genera.** A) trap caught records pooled across all sites and years, B) trap records from POS, C) trap records from SCL, D) trap records from BPF, E) trap and net caught records pooled across all sites and years, and F) all net caught records pooled across sites and years.

Or:

A graph of different colors and sizes

Description automatically generated

**Proportional abundance and unique species** for A) trap caught records and B) net caught records. Data combine records across all sites and years.

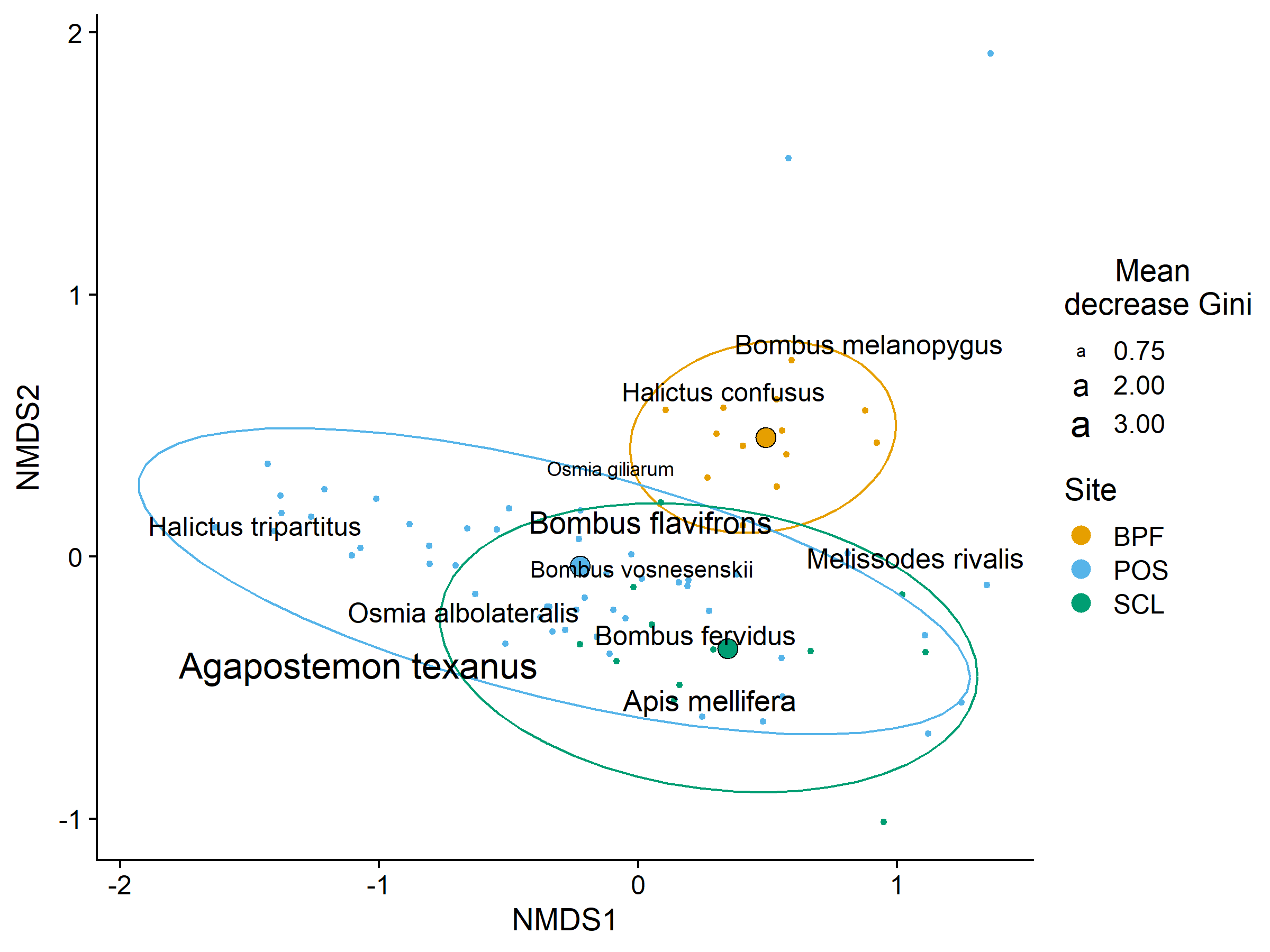
Or:

A graph of different colors and sizes

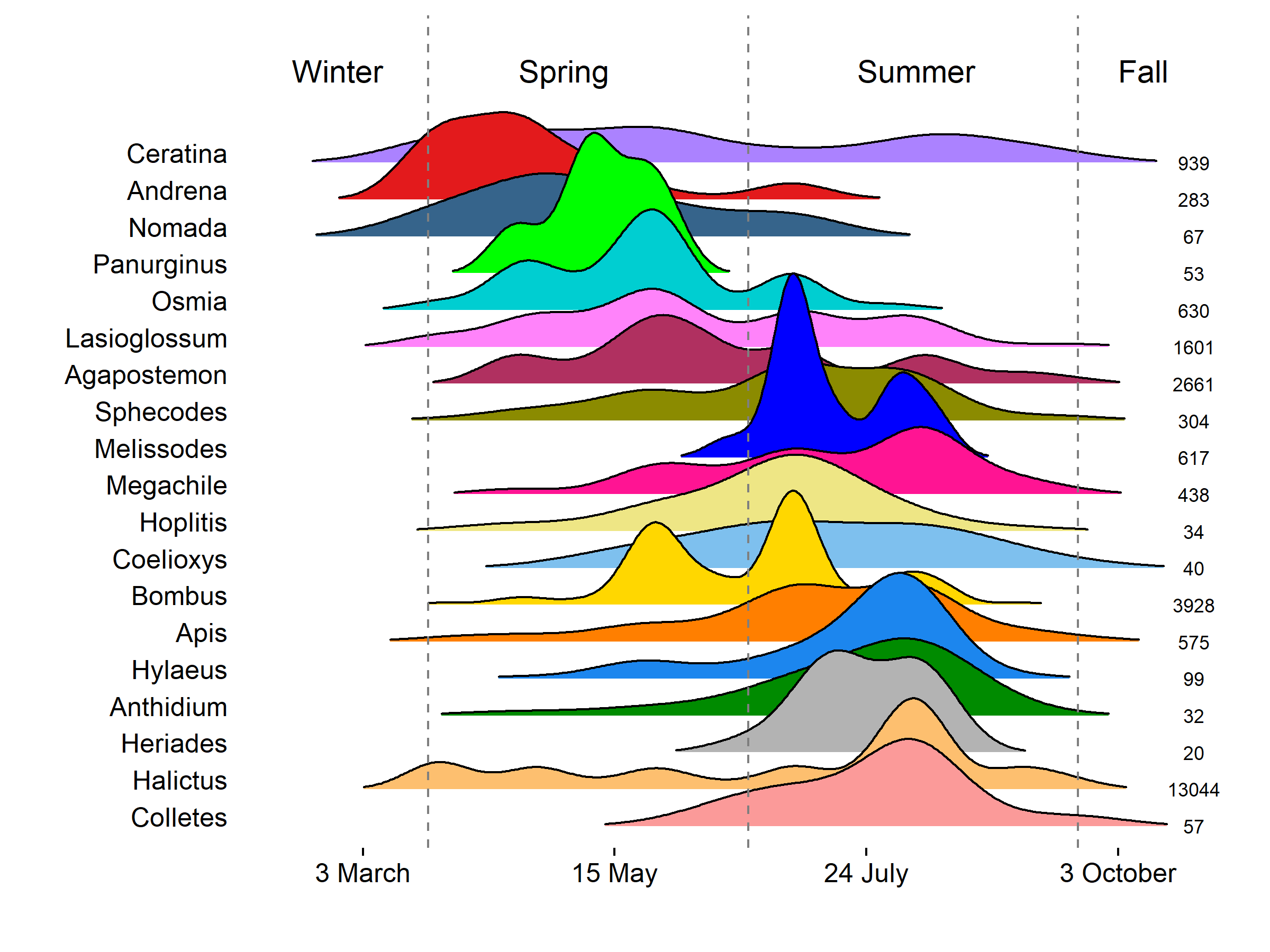
Description automatically generated

**Proportional abundance and unique species** for A) all net and trap caught records and B) net caught records only. Data combine records across all sites and years.

Species composition varied significantly across sites (PERMANOVA; *F*2 = 2.11, *P* = 0.005), with BPF hosting a statistically distinct bee community from the POS and SCL sites (Figure XX). Moreover, our random forest model classified each site by species composition with an overall out of bag error rate of 15.38%. However, while the model struggled to distinguish between POS and SCL, the BPF site was never misclassified (0.0% class error).

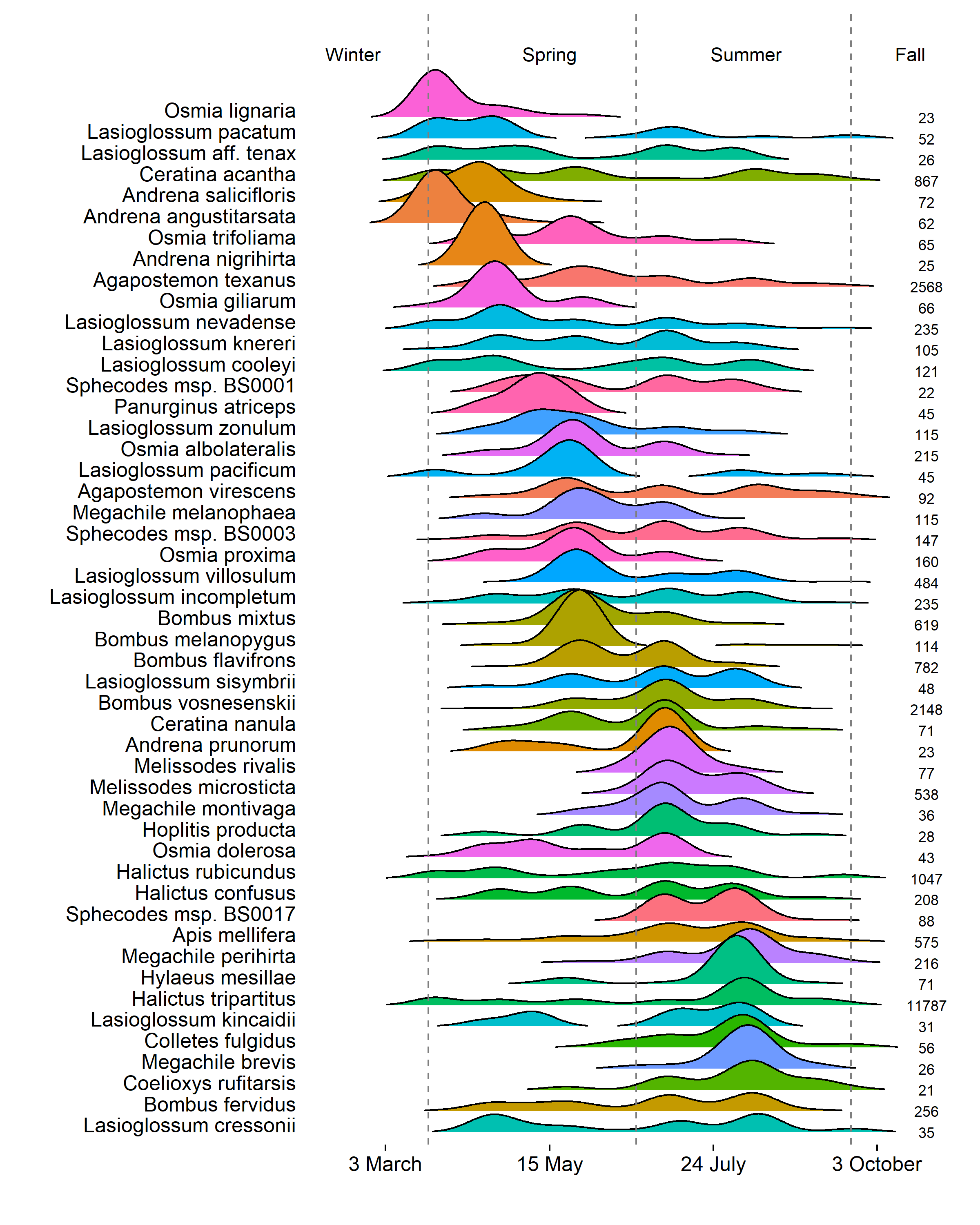


**Variation in community composition across sites.** Bee species are plotted on the first two axes of a three-dimensional non-metric multidimensional scaling ordination of the 69 combinations of station (subsite) and year, across the three sites. Small points are the individual station/year combinations. Large points are the centroids of the three sites. Ellipses are 95% confidence intervals around the site centroids. Bee species shown are the most representative (top 10th percentile of a random forest analysis) of the compositional differences among sites. Text size of the labels is proportional to variable importance score (mean decrease in Gini score).

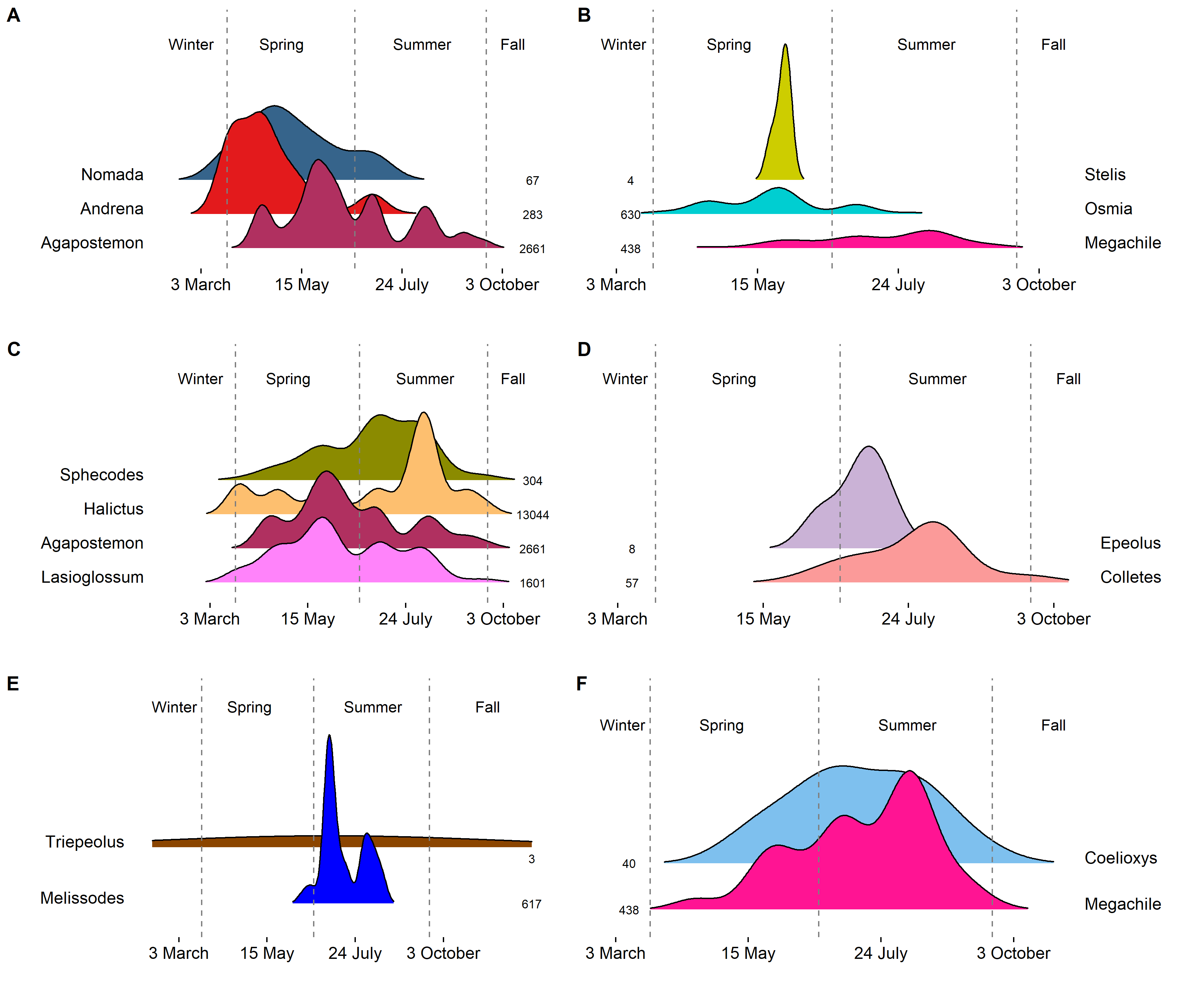


**Genus-level seasonal distributions.**Density is estimated at the genus level with Scott’s method for genera presumed univoltine, while biased cross validation was used for genera presumed multivoltine. Sample sizes displayed on the right are the total number of records for each genus. Only genera with sample sizes 20 are shown. Vertical dashed lines correspond to 21 March, 21 June, and 21 September.

Scott, D. W. (1992) *Multivariate Density Estimation: Theory, Practice, and Visualization.* New York: Wiley.

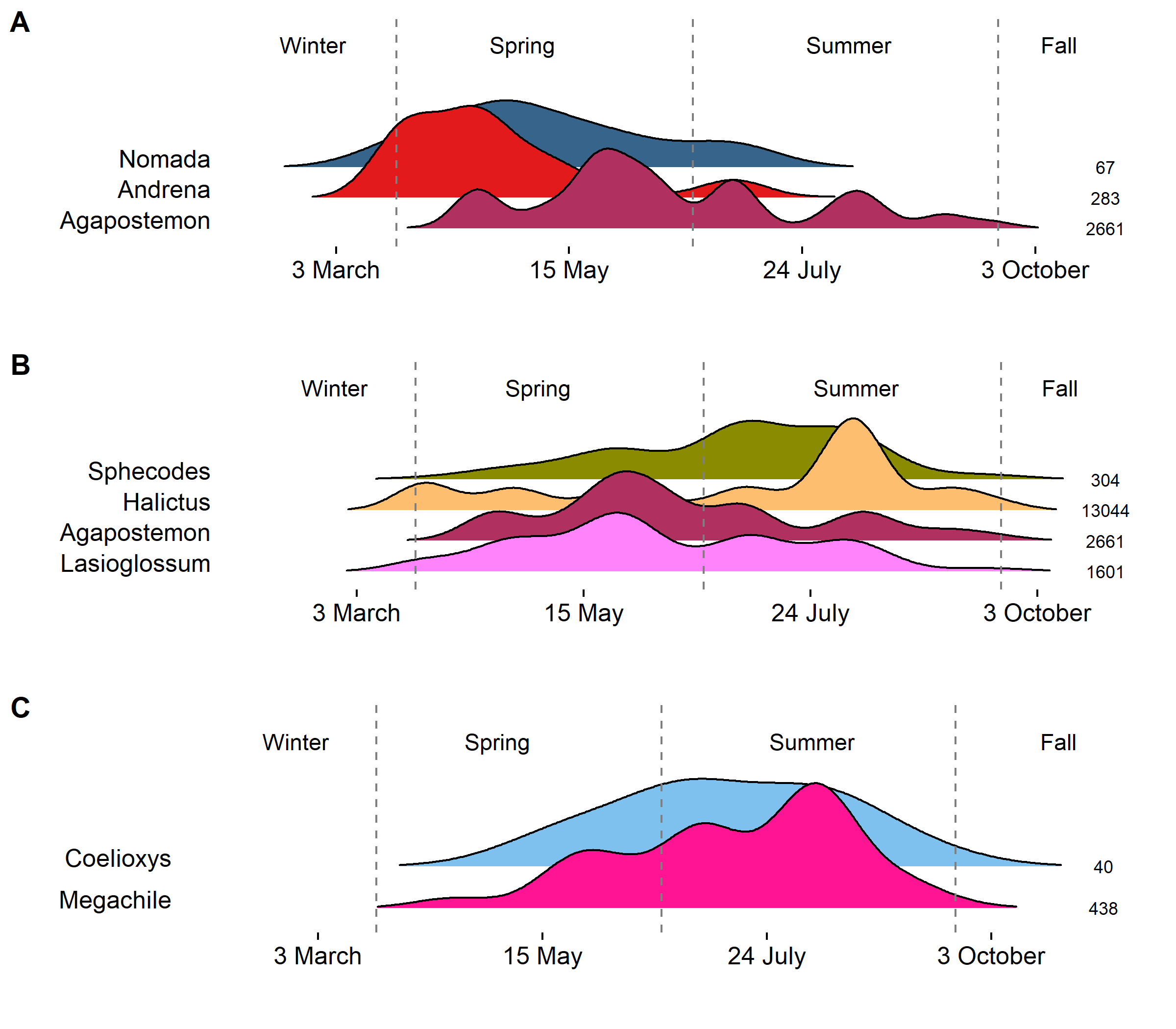


**Species-level seasonal distributions.**Density is estimated uniformly across all species with Silverman’s method. Sample sizes displayed on the right are the total number of records for each species. Species displayed are those for which sample sizes were [\ge](https://camo.githubusercontent.com/c2d96728cd1cda8c7998418bbaf71dacc073bf4fbad71ceb03065377d8251e2e/68747470733a2f2f6c617465782e636f6465636f67732e636f6d2f706e672e6c617465783f2535436765) 20. Vertical dashed lines correspond to 21 March, 21 June, and 21 September.



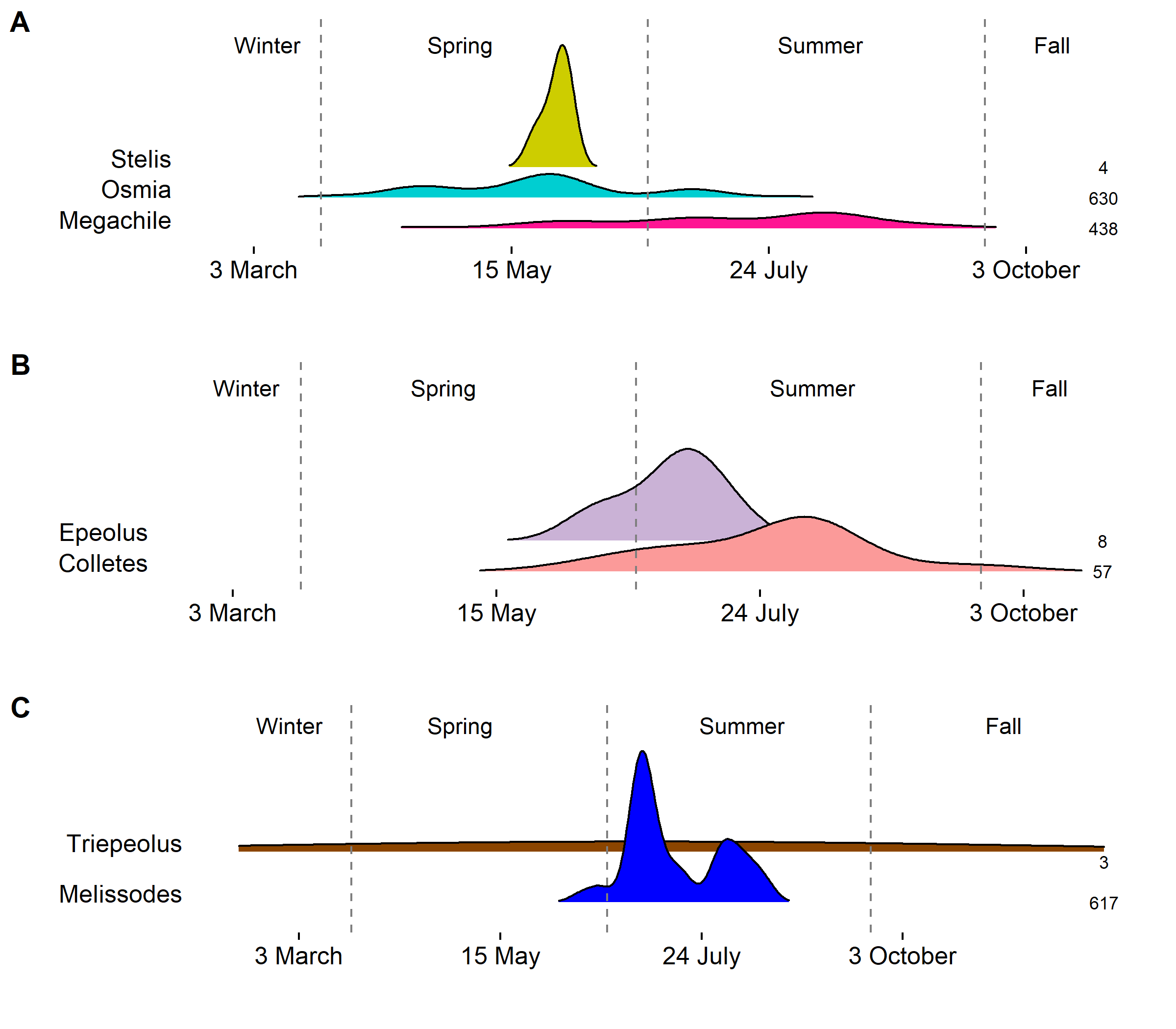
**Genus-level seasonal distributions for the parasites:** A) Nomada, B) Stelis, C) Sphecodes, D) Epeolus, E) Triepeolus, and F) Coelioxys. Beneath each parasite genera are the presumed host genera. Density is estimated at the genus level with Scott’s method for genera presumed univoltine, while biased cross validation was used for genera presumed multivoltine. Sample sizes displayed on the right (plots A, C, E) and left (plots B, D, F) are the total number of records for each genus. Vertical dashed lines correspond to 21 March, 21 June, and 21 September.

Or:



**Genus-level seasonal distributions** for the parasites: A) Nomada, B) Sphecodes, and C) Coelioxys. Beneath each parasite genera are the presumed host genera. Density is estimated at the genus level with Scott’s method for genera presumed univoltine, while biased cross validation was used for genera presumed multivoltine. Sample sizes displayed on the right are the total number of records for each genus. Vertical dashed lines correspond to 21 March, 21 June, and 21 September.

And supplement:



**Genus-level seasonal distributions for the parasites:** A) Stelis, B) Epeolus, and C) Triepeolus. Beneath each parasite genera are the presumed host genera. Density is estimated at the genus level with Scott’s method for genera presumed univoltine, while biased cross validation was used for genera presumed multivoltine. Sample sizes displayed on the right are the total number of records for each genus. Vertical dashed lines correspond to 21 March, 21 June, and 21 September. These parasites have low sample sizes limiting distributional estimation.

……………………………………….OLD VERSION…………………………………………..

*Basic results*

Number of specimens overall

Pinned vs alcohol

*Analytical results*

Abundance, Richness, Diversity: Total, by site (Table?)

Accumulation curve: Total, POS (Fig.)

Diversity by family

Spp. accumulation curve (overall)

Estimated diversity overall, by site

Most commonly collected species (10?) by numerosity, date, geography (=most pickled)

Least commonly collected species

Proportion of parasitic species

Non-native/adventive/introduced species

New regional records

DISCUSSION

Overview of results in context of urbanized habitats. Prendergast et al 2022

Approximation of richness, what numbers tell us

family diversity breakdown, Comparision to other studies

Relative diversity between sites (BPF, POS, SCL)

Why so many *H. tripartitus*? Why no males?

The preponderance of *Andrena*, *Osmia*, (“*Evalaeus/Hemihalictus*” complex) comparison to other areas, habitats; mention of recent taxonomic work on latter group.

Usefulness as baseline

Performance of trapping unit

Conservation Implications: habitat complexity, “restoration”, assessment-based strategies

(Comparison of trap performance or lack thereof.)

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The surviving authors would like to recognize our colleague and junior author, Robert Redmond, now deceased, for his creative vision and scientific facilitation, without which this work would not have been possible.

TABLES

Overall species list\*

Ten most common species by 3 criteria\*\*

\*species, family, date range of collections, sites at which collected, parasite?, non-native?, range record (State, County)

\*\*10 most common: species, overall numbers collected & proportion of males; species, date range by day of month/species; species, sites at which collected; species, stations within sites?

FIGURES

Accumulation curve overall

POS accumulation curve (finer grain, longer scale)

Diversity overlap diagram, 3 sites

Family composition overall (pie diagram)

Family composition by site (pie diagram)

SUPPLEMENTAL FIGURES

photograph of typical trap placement\*

photograph of stand\*

photograph of backpack sprayer\*

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SUPPLEMENT

Photo of WP backpack bowl filler & trap array.